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Exome Results & Raw Data Summary

Generated on: 4/12/2012

Congratulations! Your exome has been sequenced and your data is ready for you to download. We have also included this overview of your data to get you started on your exome exploration. Here are a few important points about your exome data:

- Two types of files are available for download: 1) the aligned sequencing reads in BAM format, 2) a file containing variant calls (VCF file).
- The raw data VCF file is a preliminary draft of your exome. Our ability to call variants, especially indels, is greatly improved with each additional exome added to our database. Moreover we will build upon this protocol to include additional steps such as custom treatment of the sex chromosomes. To this end we will update your VCF file at the end of the pilot. We will contact you when this data is available.

Your exome at a glance:

[Your exome in numbers](#)

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The Exome Service is a pilot project, and this report contains preliminary data only. 23andMe does not represent that all of this information is accurate. **In this report we have used 1000 Genome Project data to report frequencies of variants to determine how common or rare a particular variant is.** We have also only provided information about a subset of the many gene-disrupting variants present in the human genome, in a chosen set of genes. Sequencing was performed such that the total number of bases read was at least 80X the size of the exome. As described in the Exome Terms of Use, 23andMe will not be providing the reports and explanations that 23andMe typically provides to customers with respect to their genotyping results for this data. 23andMe Services are for research, informational, and educational use only. We do not provide medical advice. Please keep in mind that genetic information you share with others could be used against your interests.

Your exome in numbers

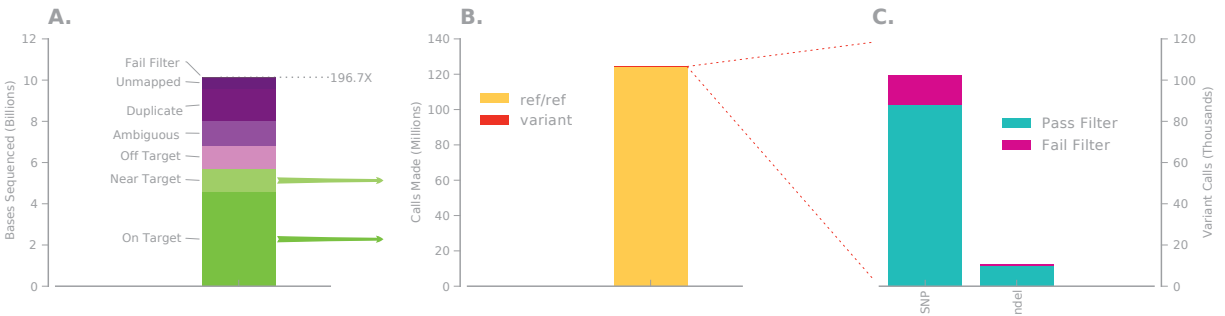


Figure 1: Getting from raw reads to called variants. A) The number of bases obtained by sequencing your exome. The top line indicates total coverage. B) Total number of called bases in your exome. The vast majority are the same as the reference genome. C) An expansion of the small sliver of variants depicted in B. These are the variants present in your VCF file.

Welcome to your exome. Your exome is the 50 million DNA bases of your genome containing the information necessary to encode all your proteins. Your exome data consists of two parts, the raw data (both aligned and unaligned Illumina reads, fig1A) and a draft of the variants present in your exome (fig1C). While this draft is provisional and we will be improving upon it, we wanted to allow you to dig in to your exome as soon as possible so you can tell us what you think is important and should be included.

To create the first draft of your exome we implemented the Broad Institute's "Best Practice" protocol for exome sequencing analysis. You can read a detailed description of it [here](#) (for brief summary see [Appendix](#)).

Characterizing your variants

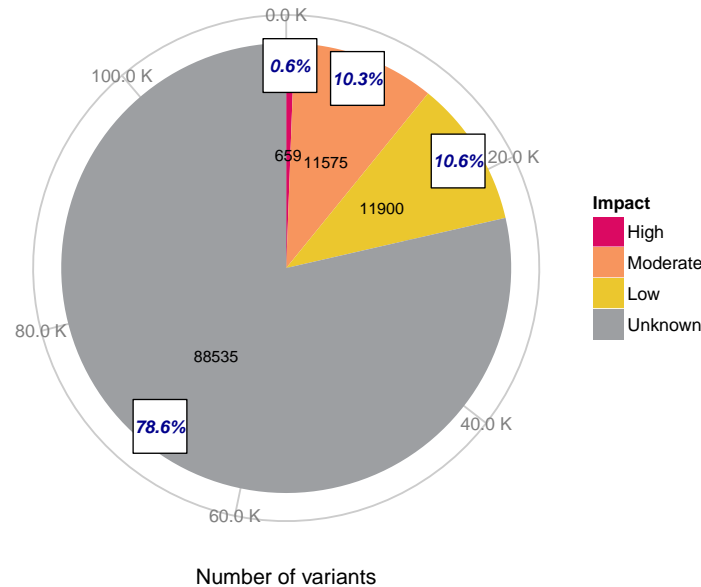


Figure 2: Predicting impact of variants on gene function. An overview of your variants and their predicted impact on gene function.

The variants in your VCF file are the positions in your genome that differ from the reference genome. Most of these variants are likely to be functionally neutral and unlikely to cause any severe disorders. Pinpointing genuine disease mutations is still challenging and we used a number of software tools to identify those that may be functionally important. We estimated the impact a variant has on gene function based on the severity of its effect on the gene product:

High impact:

Frame shift Insertion or deletion of bases, not multiple of 3.

Splice site Variant at the 'splicing site' may disrupt the consensus splicing site sequence.

Stop gain Premature termination of peptides, which would disable protein function.

Start loss Loss of the start codon.

Stop loss Loss of the stop codon.

Moderate impact:

Nonsynonymous substitution Non-conservative change altering an amino acid in a protein.

Codon insertion or deletion Insertion or deletion of bases, multiple of 3.

Low impact:

Synonymous substitution Variant that does not alter the amino acid sequence due to codon degeneracy.

Start gain Variant resulting in the gain of a start codon.

Synonymous stop Variant changing one stop codon into another.

Unknown impact: Variants unlikely to affect gene products.

How rare are your variants?

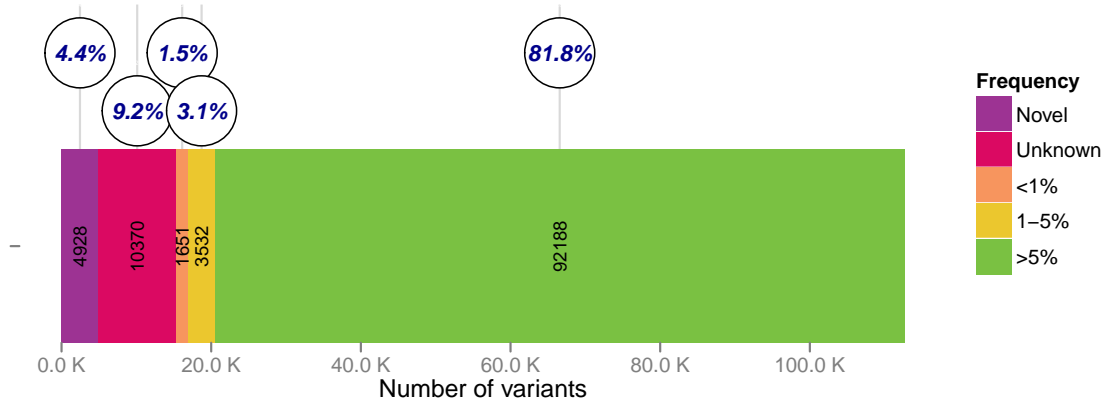


Figure 3: Variant frequencies. The allele frequencies of the variants in your exome. Unknown: allele is present in a public database but no frequency data was available.

One of the advantages of exome sequencing is that we can detect sequence variants that are unique to you! By comparing your variants to all those that have been discovered so far, we can divide your variants into the following categories:

- **novel** variant hasn't been observed in current public sequence databases
- **unknown** variant has been observed in public databases but allelic frequency has not been calculated and therefore is not available
- **rare** variant with allelic frequency $<1\%$
- **somewhat rare** variant with frequency 1-5%
- **common** frequency of the variant is greater than 5%

One of the most comprehensive human variation public datasets is maintained by the 1000 Genomes Project. We use 1000 Genomes Project data (project release: 08-26-2011) to report frequencies of alleles found in your exome, including reporting if it is absent from the public database (*i.e.* a novel variant).

Filtering your variants

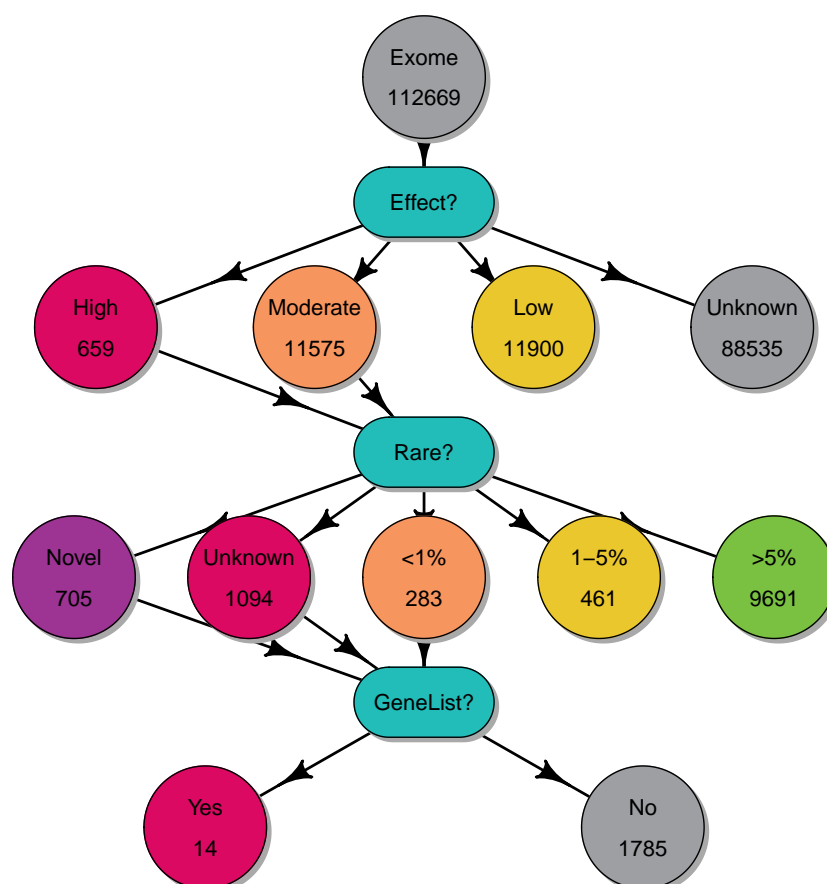


Figure 4: Variant filtering decision tree. A graphical representation of the filtering process that was used to generate your short list of variants of interest.

Most sequence variants in your exome are likely to be neutral and do not cause any severe disorders. A filtering process is often undertaken to prioritize variants discovered through sequencing. To identify potentially interesting and relevant variants with potential functional effects (contributing to disease and other phenotypes of interest) we used three consecutive filters, depicted in the figure above: (1) effect of the variant on the gene product; (2) allele frequency of the variant; (3) location of the variant in one of 592 genes involved in Mendelian disorders (at this point we also exclude indels and variants on the sex chromosomes).

We hope you find this initial list of variants interesting and that it will help you in your journey through your exome. This short list of variants only scratches the surface of what your genome contains and is just the beginning of where your data can take you. Have fun!

List of selected variants

Gene: [USH2A](#) Your genotype: **C/T** Location: chr1:215844373

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

IKGenomes: 0.00320

dbSNP: [rs45549044](#)

Genotype quality: 99

Coverage depth: 137

EntrezId: 7399

EnsemblId: [ENSG00000042781](#)

UniProt: [O75445](#)

OMIM: 608400

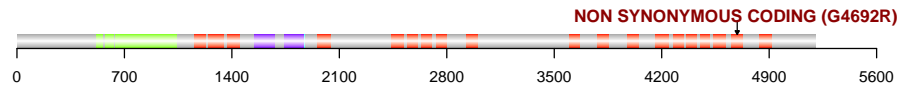
Gene Description: Usher syndrome 2A (autosomal recessive, mild)

Transcript: [ENST00000307340](#)

AA change: G4692R

PFAM (or SMART) domains for gene USH2A, transcript ENST00000307340:

- PF00053: EGF_laminin
- PF00041: FN_III
- PF00054: Laminin_G_1
- PF02210: Laminin_G_2



Gene: [LRP5](#) Your genotype: **C/T** Location: chr11:68213989

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

IKGenomes: 0.00610

dbSNP: [rs1127291](#)

Genotype quality: 99

Coverage depth: 55

EntrezId: 4041

EnsemblId: [ENSG00000162337](#)

UniProt: [O75197](#)

OMIM: 603506

Gene Description: low density lipoprotein receptor-related protein 5

Transcript: [ENST00000294304](#)

AA change: A1525V

PFAM (or SMART) domains for gene LRP5, transcript ENST00000294304:

- PF00058: LDLR_classB_rpt
- PF00057: LDrepeatLR_classA_rpt



Gene: **VPS13A** Your genotype: **A/G** Location: chr9:79936140

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 5e-04

dbSNP: [rs144290291](#)

Genotype quality: 99

Coverage depth: 49

EntrezId: 23230

EnsemblId: [ENSG00000197969](#)

UniProt: [Q96RL7](#)

OMIM: 605978

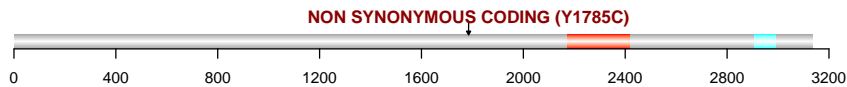
Gene Description: vacuolar protein sorting 13 homolog A (S. cerevisiae)

Transcript: [ENST00000376636](#)

AA change: Y1785C

PFAM (or SMART) domains for gene VPS13A, transcript ENST00000376636:

- PF06650: VPSAP
- PF09333: Autophagy-rel_C



Gene: **F5** Your genotype: **T/A** Location: chr1:169511585

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 0.00870

dbSNP: [rs9332695](#)

Genotype quality: 99

Coverage depth: 153

EntrezId: 2153

EnsemblId: [ENSG00000198734](#)

UniProt: [P12259](#)

OMIM: 612309

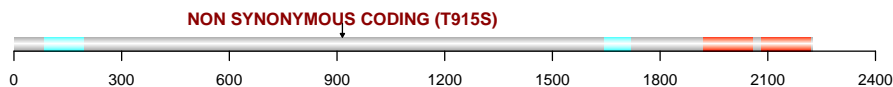
Gene Description: coagulation factor V (proaccelerin, labile factor)

Transcript: [ENST00000367797](#)

AA change: T915S

PFAM (or SMART) domains for gene F5, transcript ENST00000367797:

- PF07732: Cu-oxidase_3
- PF00754: Coagulation_factor_5/8-type_C



Gene: [CNGB3](#) Your genotype: [T/C](#) Location: chr8:87755776

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 0.00960

dbSNP: [rs35807406](#)

Genotype quality: 99

Coverage depth: 76

EntrezId: 54714

EnsemblId: [ENSG00000170289](#)

UniProt: [Q9NQW8](#)

OMIM: 605080

Gene Description: cyclic nucleotide gated channel beta 3

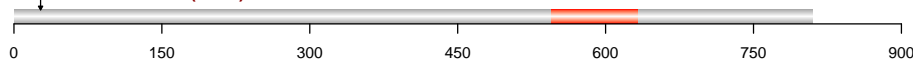
Transcript: [ENST00000320005](#)

AA change: N27S

PFAM (or SMART) domains for gene CNGB3, transcript ENST00000320005:

■ PF00027: cNMP-bd_dom

NON SYNONYMOUS CODING (N27S)



Gene: [ATR](#) Your genotype: [A/G](#) Location: chr3:142212065

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 9e-04

dbSNP: [rs150339560](#)

Genotype quality: 99

Coverage depth: 93

EntrezId: 545

EnsemblId: [ENSG00000175054](#)

UniProt: [Q13535](#)

OMIM: 601215

Gene Description: ataxia telangiectasia and Rad3 related

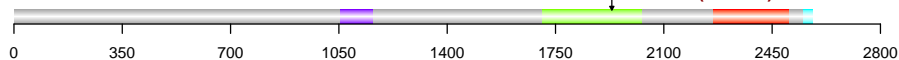
Transcript: [ENST00000383101](#)

AA change: M1932T

PFAM (or SMART) domains for gene ATR, transcript ENST00000383101:

■ PF08064: UME
■ PF02259: PIK-rel_kinase_FAT
■ PF00454: PI3/4_kinase_cat
■ PF02260: FATC

NON SYNONYMOUS CODING (M1932T)



Gene: [DPYD](#) Your genotype: **C/C** Location: chr1:98144726

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 0.00500

dbSNP: [rs45589337](#)

Genotype quality: 51.14

Coverage depth: 24

EntrezId: 1806

EnsemblId: [ENSG00000188641](#)

UniProt: [Q12882](#)

OMIM: 612779

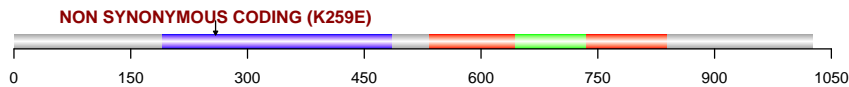
Gene Description: dihydropyrimidine dehydrogenase

Transcript: [ENST00000370192](#)

AA change: K259E

PFAM (or SMART) domains for gene [DPYD](#), transcript [ENST00000370192](#):

- PF07992: Pyr_nucl-diS_OxRdtase_FAD/NAD
- PF01180: Dihydroorotate_DH_1_2
- PF01207: tRNA_hU_synthase



Gene: [D2HGDH](#) Your genotype: **G/A** Location: chr2:242695399

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 0.00340

dbSNP: [rs146578303](#)

Genotype quality: 99

Coverage depth: 55

EntrezId: 728294

EnsemblId: [ENSG00000180902](#)

UniProt: [Q8N465](#)

OMIM: 609186

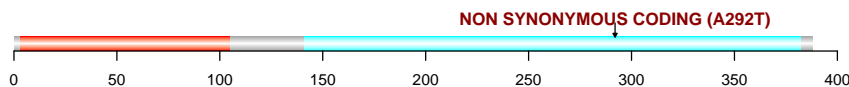
Gene Description: D-2-hydroxyglutarate dehydrogenase

Transcript: [ENST00000403782](#)

AA change: A292T

PFAM (or SMART) domains for gene [D2HGDH](#), transcript [ENST00000403782](#):

- PF01565: Oxid_FAD_bind_N
- PF02913: FAD-linked_oxidase_C



Gene: [INSR](#) Your genotype: [C/T](#) Location: chr19:7125518

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 0.00360

dbSNP: [rs1799816](#)

Genotype quality: 99

Coverage depth: 77

EntrezId: 3643

EnsemblId: [ENSG00000171105](#)

UniProt: [P06213](#)

OMIM: [147670](#)

Gene Description: insulin receptor

Transcript: [ENST00000341500](#)

AA change: V1000M

PFAM (or SMART) domains for gene INSR, transcript ENST00000341500:

- PF01030: EGF_rcpt_L
- PF00757: Furin-like_Cys-rich_dom
- PF00041: FN_III
- PF07714: Ser-Thr/Tyr_kinase
- PF00069: Se/Thr_kinase-like_dom



Gene: [VPS13A](#) Your genotype: [C/T](#) Location: chr9:79938036

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 0.00510

dbSNP: [rs149694033](#)

Genotype quality: 99

Coverage depth: 43

EntrezId: 23230

EnsemblId: [ENSG00000197969](#)

UniProt: [Q96RL7](#)

OMIM: [605978](#)

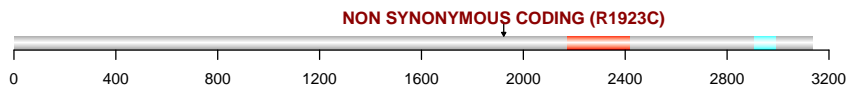
Gene Description: vacuolar protein sorting 13 homolog A (S. cerevisiae)

Transcript: [ENST00000376636](#)

AA change: R1923C

PFAM (or SMART) domains for gene VPS13A, transcript ENST00000376636:

- PF06650: VPSAP
- PF09333: Autophagy-rel_C



Gene: [TTN](#) Your genotype: **T/G** Location: chr2:179477267

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 0.00830

dbSNP: [rs36043230](#)

Genotype quality: 99

Coverage depth: 166

EntrezId: 7273

EnsemblId: [ENSG00000155657](#)

UniProt: [Q8WZ42](#)

OMIM: 188840

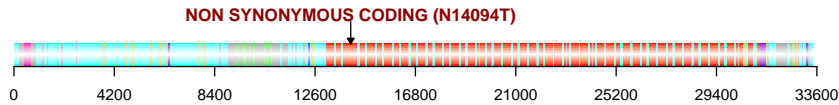
Gene Description: titin

Transcript: [ENST00000342992](#)

AA change: N14094T

PFAM (or SMART) domains for gene TTN, transcript ENST00000342992:

- PF07679: Ig_I-set
- PF09042: Titin_Z
- PF07686: Ig_V-set
- PF00047: Immunoglobulin
- PF02818: PPAK_motif
- PF00041: FN_III
- PF00069: Se/Thr_kinase-like_dom
- PF07714: Ser-Thr/Tyr_kinase



Gene: [PKD1](#) Your genotype: **G/A** Location: chr16:2161666

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 0.00850

dbSNP: [rs146887330](#)

Genotype quality: 83.6

Coverage depth: 8

EntrezId: 5310

EnsemblId: [ENSG00000008710](#)

UniProt: [P98161](#)

OMIM: 601313

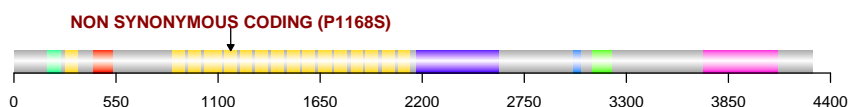
Gene Description: polycystic kidney disease 1 (autosomal dominant)

Transcript: [ENST00000262304](#)

AA change: P1168S

PFAM (or SMART) domains for gene PKD1, transcript ENST00000262304:

- PF01822: WSC_carb-bd
- PF00801: PKD_dom
- PF00059: C-type_lectin
- PF02010: PKD/REJ-like
- PF01825: GPS_dom
- PF01477: LipOase_LH2
- PF08016: PKD1_2_channel



Gene: [RELN](#) Your genotype: [C/T](#) Location: chr7:103234202

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 0.00780

dbSNP: [rs55689103](#)

Genotype quality: 99

Coverage depth: 114

EntrezId: 5649

EnsemblId: [ENSG00000189056](#)

UniProt: [P78509](#)

OMIM: [600514](#)

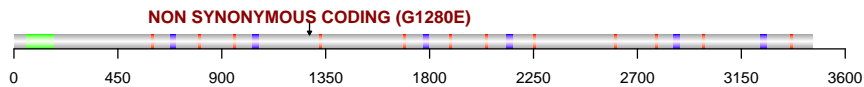
Gene Description: reelin

Transcript: [ENST00000343529](#)

AA change: G1280E

PFAM (or SMART) domains for gene RELN, transcript ENST00000343529:

- PF02014: Reeler_dom
- PF02012: BNR_rpt
- PF07974: EGF_extracell



Gene: [GPR98](#) Your genotype: [G/A](#) Location: chr5:89948189

Effect: NON SYNONYMOUS CODING **Impact:** MODERATE

1KGenomes: 8e-04

dbSNP: NA

Genotype quality: 99

Coverage depth: 213

EntrezId: 84059

EnsemblId: [ENSG00000164199](#)

UniProt: [Q8WXXG9](#)

OMIM: [602851](#)

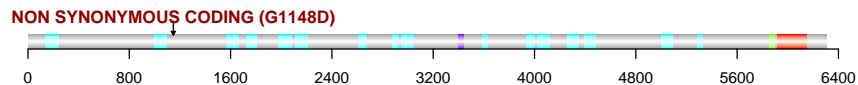
Gene Description: G protein-coupled receptor 98

Transcript: [ENST00000296619](#)

AA change: G1148D

PFAM (or SMART) domains for gene GPR98, transcript ENST00000296619:

- PF03160: Calx_beta
- PF03736: EPTP
- PF01825: GPS_dom
- PF00002: GPCR_2_secretin-like



Appendix

To create the first draft of your exome we implemented the Broad Institute's "Best Practice" protocol for exome sequencing analysis. You can read a detailed description of it [here](#), however a brief summary of it follows:

1. We took your raw reads and aligned them against the reference genome (these are the alignments available in the BAM file of the encrypted download).
2. We used these alignments to identify probable contamination (unaligned reads) and artifacts of sample preparation (PCR duplicates) which are then removed from subsequent steps.
3. From this point on we focus on the reads that align either to one of the exons or within the regions 250 bases up and downstream of it.
4. To improve the quality of the alignments we carry out a more accurate alignment of the reads that overlap known indels or are likely to contain indels themselves.
5. We also recalibrate the base quality scores of the reads to bring them in line with the empirically-determined values.
6. Using these realigned+recalibrated reads we generate allele calls at every position with enough high-quality data and filter out those that are homozygous for the allele present in the reference genome (the vast majority of these are at such a high frequency in the population they're unlikely to be interesting). The remaining SNP and indel calls (variants) are the ones available in the VCF file that you downloaded.
7. As yet no sequencing technology is 100% accurate and the highly duplicated nature of the human genome makes variant calling a challenging task. Consequently, a small proportion of the variant calls in your VCF are likely to be incorrect. To reduce this proportion we applied the filters recommended by the Broad Institute to remove technical artifacts. Variants that pass all filters are marked in your VCF file with a PASS. As the exome pilot progresses and we gather more data we will be able to use more advanced techniques identify potential errors and improve the quality of your exome.